



Paper Accepted^{*}

ISSN Online 2406-0895

Original Article / Оригинални рад

Ognjenka Janković^{1,†}, Smiljana Paraš², Ljiljana Tadić-Latinović³, Renata Josipović¹, Vukoman Jokanović⁴, Slavoljub Živković⁵

Biocompatibility nanostructured biomaterials based on calcium aluminate

Биокомпатибилност наноструктурних биоматеријала

на бази калцијум-алумината

¹University of Banja Luka, Faculty o Medicine, Department of Stomatology, Banjaluka, Bosnia and Herzegovina; ²University of Banja Luka, Faculty of Science and Mathematics, Department of Zoology, Banjaluka, Bosnia and Herzegovina;

³University of Banja Luka, Faculty of Medicine, Institute of Pathology, Clinical Center, Banjaluka, Bosnia and Herzegovina;

⁴Institute of Nuclear Sciences, Vinca, Laboratory for Atomic Physics, Belgrade, Serbia;

⁵ University of Belgrade, Faculty of Dental Medicine, Department of Restorative Dentistry and Endodontics, Belgrade, Serbia

Received: December 11, 2017 Revised: March 26, 2018 Accepted: March 29, 2018 Online First: April 3, 2018 DOI: https://doi.org/10.2298/SARH171211030J

When the final article is assigned to volumes/issues of the journal, the Article in Press version will be removed and the final version will appear in the associated published volumes/issues of the journal. The date the article was made available online first will be carried over.

[†] **Correspondence to:** Ognjenka JANKOVIĆ Faculty o Medicine, Department of Stomatology, 78000 Banjaluka, Bosnia and Herzegovina, E-mail: **ognjenka.olja@gmail.com**

^{*} Accepted papers are articles in press that have gone through due peer review process and have been accepted for publication by the Editorial Board of the *Serbian Archives of Medicine*. They have not yet been copy edited and/or formatted in the publication house style, and the text may be changed before the final publication.

Although accepted papers do not yet have all the accompanying bibliographic details available, they can already be cited using the year of online publication and the DOI, as follows: the author's last name and initial of the first name, article title, journal title, online first publication month and year, and the DOI; e.g.: Petrović P, Jovanović J. The title of the article. Srp Arh Celok Lek. Online First, February 2017.

Biocompatibility nanostructured biomaterials based on calcium aluminate Биокомпатибилност наноструктурних биоматеријала на бази калцијум-алумината

SUMMARY

Introduction/Objective The aim of this paper was to verify the biocompatibility of the newly synthesized nanostructured material based on calcium aluminate after implantation into the subcutaneous tissue of rats. **Methods** The study included 18 rats aged from 10 to 11 weeks, divided into two experimental groups (n=9). In all animals, incision took place on the back and two pockets of 15 mm in depth were made, in which sterile polyethylene tubes with test materials (*ALBO*-CA, *ALBO*-CSHA and MTA control group) were implemented. Six rats of each group were sacrificed in three observational periods (7, 15, 30 days). Patohystological analysis included: inflammation, bleeding, fibrous capsule, and tissue integrity around the implanted material.

Results After 7 days of threatment, *ALBO*-CA and *ALBO*-CSHA showed better tissue response compared to MTA with a statistically significant difference in inflammation intensity (p=0.2781). The difference in vascular congestion and thickness of the fibrous capsule after implantation of *ALBO*-CA material compared to MTA was also statistically significant (p=0.5567). At the end of the 30-day evaluation period, an identical inflammatory response of connective tissue at the site of implantation *ALBO*-CA, *ALBO*-CSHA and MTA (score 0.7) was recorded. The formation of thick or moderately thick fibrous capsule was found to be thickest in *ALBO*-CA (grade 3.7). There were no statistically significant differences between the parameters analyzed after 30 days.

Conclusion Newly-synthesized *ALBO*-CA showed a satisfactory tissue response and confirmed biocompatibility after implantation in subcutaneous tissue of rats.

Keywords: nanomaterials; calcium aluminates; calcium silicates; tissue reaction

Сажетак

Увод/Циљ Циљ овог рада је био да се провери биокомпатибилност новосинтетисаног наноструктурног материјала на бази калцијум-алумината након имплантације у поткожно ткиво пацова. Методе У истраживање је укључено 18 пацова старости 10-11 седмица који су подељени у две експерименталне групе (n=9). Код свих животиња је урађена инцизија на леђима и формирана два цепа дубине 15 мм у које су аплициране стерилне полиетиленске тубице са тестним материјалима (ALBO-CA, ALBO-CSHA и контрола MTA). По шест пацова сваке групе је жртвовано у три опсервациона периода (7, 15, 30 дана). Патохистолошки су анализирани: инфламација, крварење, фиброзна капсула и интегритет ткива око имплантираног материјала. Резултати Након седам дана ALBO-CA и ALBO-

CSHA су показали бољи ткивни одговор у односу на МТА са статистички значајном разликом у интензитету инфламације (р=0,2781). Разлика у васкуларној конгестији и дебљини фиброзне капсуле након имплантације материјала ALBO-CA у односу на МТА је такође била статистички значајна (*p*=0,5567). На крају евалуационог периода од 30 дана је забележен идентичан инфламаторни одговор растреситог везивног ткива на мјесту имплантације ALBO-CA, ALBO-CSHA и МТА (оцена 0,7). Констатовано је формирање дебеле или умерено дебеле фиброзне капсуле која је била најдебља код АLBO-СА (оцена 3,7). Статистички значајне разлике између анализираних параметара након 30 дана није било. Закључак Новосинтетисани ALBO-CA је показао задовољавајућу ткивну реакцију и потврдио биокомпатибилност имплантације након V поткожно ткиво пацова.

Кључне речи: наноматеријали; калцијумалуминати; калцијум-силикати, ткивна реакција

INTRODUCTION

In recent years, tendencies of scientific research focused on the discovery of new biomaterials that would through changes in composition and improved characteristics overcome the shortcomings of the existing ones. In contact with tissue, biomaterials must not exhibit potential cytotoxic, genotoxic, mutagenic or allergic reactions [1].

Biocompatibility tests are carried out *in vitro* and *in vivo* conditions and include: testing of material in laboratory conditions on cell tissues or organs cultures, animal experiments and clinical tests on humans. The efficiency of animal tests is more important than *in vitro* research. These tests

are carried out by using subcutaneous implantation, allergic tests and tests of acute and chronic systemic toxicity of dental materials [2].

Calcium aluminate cements (CAC) were developed at the beginning of the 20th century as chemically durable materials from Portland cement. Their hydraulic calcium phase contains from 30% to 50% of aluminum, while in Portland cement, aluminum is less than 5%. [3-5].

More intense studies of the potential biomedical application of calcium aluminate cements have been carried out a decade ago.

Attempts of calcium aluminates usage in dentistry for direct restorative fillings on lateral teeth (DoxaDent; Doxa AB, Uppsala, Sweden, Scandinavia) (Ceramir C&B, Doxa AB), during a three-year clinical monitoring did not indicate clinical success, due to inferior mechanical properties and handling difficulties [6,7].

Calcium aluminate in addition with X-ray contrast agent, mixed with water or saline solution 0,9 %, resulted in endodontic cement of similar indications as commercially available tricalcium silicate cements (EndoBinder, Binderweare, Sao Carlos, SP, Brazil). EndoBinder was developed with the intention of preserving the positive properties and clinical application of MTA, but without its negative properties. According to the results of several recent studies, EndoBinder showed appropriate biological and physico-chemical characteristics, good antimicrobial activity against *Enterococus fecalis, Staphylococcus aureus*, and *Candide albicans*, and after the implantation into subcutaneous tissue of rats caused a minor inflammatory reaction compared to MTA [8-11] or comparable to MTA [4]. During the repair of bone defects, EndoBinder showed similar behavior as MTA, and stimulated complete bone recovery after 3 months [12].

Quick-set (Primus Consulting, Bradenton, FL), calcium aluminate cement, confirmed the dentinogenic potential as WMTA (Dentsply Tulsa Dental Specialties, Tulsa, OK) in *in vitro* conditions on the MDPC-23 odontoblast cell line [13] *in vivo* conditions in pulpotomy [14].

Attempts of combining calcium aluminate cements (CAC) with collagen, zirconium, zinc oxide, tricalcium phosphate and hydroxyapatite have shown great potential for application in odontology and orthopedics. Compared with commercial products for implementation in dentistry (MTA, glasionomer cement Vidrion) and orthopedics (PMMA), CAC mixtures showed greater alkaline phosphatase activity, higher cellular viability of osteoblast cells, and formation apatit on the surfaces of all CAC samples of homogeneous layer [15].

The combination of tricalcium aluminates $(Ca_3Al_2O_6)$ with tricalcium silicates (Ca_3SiO_5) resulted in better physico-chemical properties (accelerated the hydration process, reduced the setting time and improved the compressive strength), higher bioactivity and biocompatibility (stimulating effect on L929 cell growth) compared to pure tricalciumilicates (Ca_3SiO_5) [16].

Despite the fact that in last decade a emphasis has been placed on research into calcium aluminate cements, wider use of these materials in dental practice requires additional testing of a recent date.

Research on new calcium aluminates with similar physical and biological properties as well as MTA, but without its negative characteristics are still current.

Simultaneously, there is a growing influence of nanotechnology and nanomaterials in medicine.

The objective of this study was to investigate the biocompatibility of the newly synthesized nanostructured material based on calcium aluminate after implantation into the subcutaneous tissue of rats.

METHODS

Research was conducted after the approval of the Ethic Committee of the University Clinical Center in Banja Luka, No. 01-9-192.2/15, Bosnia and Herzegovina, and an experiment was realized at the Vivarium of the Faculty of Natural Sciences and Mathematics, University of Banja Luka and in the Laboratory of the Institute of Pathology, Clinical Center Banja Luka.

Tested materials

The calcium aluminate-based nanomaterial system (*ALBO*-CA) was compared to the hydroxyapatite and calcium silicate based material (*ALBO*-CSHA). White Mineral Trioxide Aggregate (MTA Angelus® Tulsa OK, USA) was used as a positive control.

The calcium aluminate system (CaO • $Al_2O_3 + CaCO_3 + Bi_2O_3$) is a mixture called *ALBO*-MCCA, obtained by mixing CaCO₃ and Bi₂O₃, and BaSO₄ with a calcium aluminate phase in the ratio of 2: 2: 1. In order to obtain calcium aluminate endodontic mixtures, it was necessary first to synthesize particular components of the mixture: calcium aluminate (CaO•Al₂O₃, CA) and calcite (CaCO₃). The mixture was finally mixed with distilled water in the 2:1 ratio of powder to water, according to the recommended protocol.

Another test material was calcium silicate with the addition of hydroxyapatite. Both of the materials were synthesized according to the method of Professor Jokanović and his associate using newtechnology, a combination of hydrothermal sol-gel method and the method of self combusting waves [17].

Design of the study

The animal model were rats of Wistar strain (18 rats, aged from 10 to 11 weeks, average weight from 190 to 280 g). During the experiment rats had free access to food and water, a 12-hour shift of light and dark, air temperature ranged from 20 to 23 °C, and humidity $60\% \pm 10\%$. Rats were divided into two experimental groups (n = 9). 6 rats (3 from both groups) were sacrificed after 7, 15 and 30 days. Subcutaneous implantation of polyethylene tubes (length 10 mm, inner diameter 1 mm) was conducted, up to half filled with tested materials (*ALBO*-CA I group, *ALBO*-CSHA II group) and half empty. An empty half of the tube was a negative control. Two tubes were placed on the back of each rat, on the right side of the tube with the tested material, and on the left side of the spinal column of

the tube with the MTA (positive control). Tubes were oriented so that the material was always turned to the head, and the empty part of the tube to the tail.

Before the surgical procedure, rats were introduced to general anesthesia (Ketamin 90 mg / kg body weight, Ketamine Hydrochloride Injection USP Rotexmedica-Germany in combination with Xylazine 5 mg / kg body weight 2% Xylazine, Cp Pharma, Bergdorf, Germany).

Preparation of the operational field was carried out, and then a blunt dissection, right and left of the spinal column, two pockets of depth about 15 mm were formed. Sterile polyethylene tubes, previously filled with freshly mixed test materials, are placed in the pockets in this way. After that, the wound was sewed. An individual knotted seam was applied.

After the operation animals were placed in one cage in the controlled environment, with controlled diet and daily professional care.

Animal health control was carried out three times a day. Six animals were sacrificed in each observation period (7, 15, 30 days). For this purpose, intravenous injection of Pentobarbitol (Pentobarbital sodium salt 100mg / ml, Sigma-Aldrich Chemie GmbH, Steinheim, Germany) was used.

Prepared subcutaneous tissue, together with tubes, was immersed in 10% formalin and delivered to laboratories for histological analysis.

For histological analysis, 4 samples were taken from each animal. After that, tissue clip fixation together with polyethylene tubes was performed in 10% buffered formalin, paraffin molding, and paraffinic dyeing (4 µm thickness) of hematoxylin eosin (HE). Analysis of the preparation was done on a light microscope (Olympus BX-51, Japan) by an experienced pathologist, who did not participate in the sampling of the material.

Histological analysis of the prepared samples was performed qualitatively and semiquantitative, and the inflammatory reaction, vascular congestion, fibrous capsule and the preservation of the integrity of the connective tissue were considered.

Statistical analysis

Dunn's post-hoc test was applied for the significance of differences between pairs with a significance level of a = 0.05 for statistical analysis of the obtained subcutaneous implantation results. Statistical analysis was done in the Minitab 16 software package (Minitab INC. State College, PA, USA).

RESULTS

Results of the histological analysis are shown in Table 1 and Figures 1–7.

After a 7-day histopathological analysis, calcium aluminate *ALBO*-CA and calcium silicate hydroxyapatite *ALBO*-CSHA a moderate inflammatory reaction was recorded (grade 3), while in MTA in 3 cases was moderate (grade 3) and in 3 cases an intensive inflammatory reaction (grade 4),

with a statistically significant difference (p=0.2781). Moderate inflammation was traced by moderate vascular congestion in *ALBO*-CA (grade 3) and *ALBO*-CSHA material. After MTA implantation,

Parameters		7 days	15 days	30 days
Inflammatory reaction	ALBO-CA*	3 ± 1.5	2 ± 1	0.666 ± 0.5
	ALBO-CSHA [†]	3 ± 1.5	2 ± 1	0.666 ± 0.5
	MTA^\ddagger	3 ± 1.414	2 ± 1	1 ± 0.5
Vascular congestion	ALBO-CA	3 ± 1.2777	2 ± 0.8888	0.6666 ± 0.5211
	ALBO-CSHA	3.333 ± 0.5773	2.6666 ± 0.5773	0.6666 ± 0.57735
	MTA	3.6667 ± 0.5773	2.6666 ± 0.5773	1 ± 0.0
Fibrous capsule	ALBO-CA	1 ± 0.2778	2.3333 ± 1.156	4 ± 1.8888
	ALBO-CSHA	0.6666 ± 0.5774	2.3333 ± 0.5775	3.3333 ± 0.57735
	MTA	0.3333 ± 0.57735	2 ± 0.0	3.3333 ± 0.57735
Preserving the integrity of the connective tissue	ALBO-CA	3 ± 0.0	2 ± 0.0	1 ± 0.4444
	ALBO-CSHA	3 ± 0.0	2 ± 0.0	0.6666 ± 0.57735
	MTA	3 ± 0.0	2 ± 0.0	1 ± 0.0

Table 1. Analyzes of inflammation, bleeding, fibrous capsules and integrity of binders of tested materials by observation periodicity.

*ALBO-CA - calcium aluminate cement

[†]ALBO-CSHA – calcium silicate cement with the addition of hydroxyapatite

[‡]MTA – mineral trioxide aggregate

equally moderate distribution (grade 3) and pronounced vascular congestion were observed. (grade 4). In all tested materials, a moderate distortion of the connective tissue structure was registered, while the fibrous capsule after the implantation of *ALBO*-CA material was minimal (grade 1), ie minimal (grade 1) or absent (grade 0) after implantation of *ALBO*-CSHA and MTA (Figure 1 and 2). The difference in vascular congestion and the thickness of the fibrous capsule after implantation of *ALBO*-CSHA.

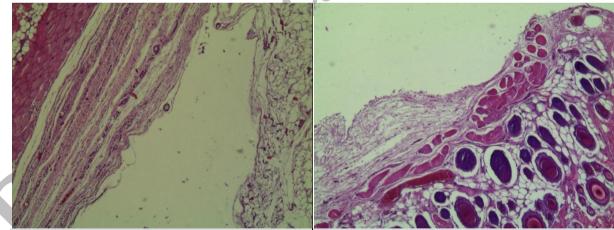
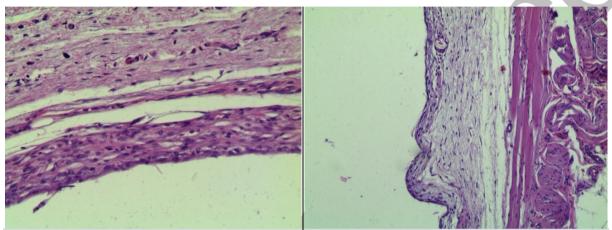


Figure 1. ALBO-CA material in the experimental Figure 2. Material ALBO-CSHA in the period of 7 days. On the microphotography, a experimental period 7 of days. In the defect (from the tube) with a very thin capsule and microphotography, the fibrous capsule is not moderate vascular congestion. present (H&E, ×200).

In the control preparations, 7 days after subcutaneous implantation, the presence of moderate intensity inflammatory reaction, and one case of pronounced inflammatory reaction in the control of the *ALBO*-CA material and the MTA control was noted. Blood vessels showed signs of moderate vascular congestion, which was more expressed in the control group of MTA. In all control preparations, there was a moderate disturbance of connective tissue structure, except in one case of

MTA control group with a slightly disrupted connective tissue structure (grade 2). The fibrous capsule was absent in all control preparations of *ALBO*-CA, and in most of the preparations of the MTA control group, except in one case. Material control of *ALBO*-CSHA recorded a gentle thin capsule in 2 cases (grade 1), and complete absence of the capsule in 1 sample (grade 0).

A pathohistological analysis after 15 days at the site of implantation of all examined materials showed a slight inflammatory reaction thin or moderately thick fibrous capsule (Figure 3 and 4), and a mild disturbance of the connective tissue structure (grade 2). Vascular congestion is characterized as mild (grade 2) in *ALBO*-CA material, or as moderate or mild to *ALBO*-CSHA and MTA, with a statistically significant difference (p=0.2974).



the Figure 4. ALBO-CSHA material in an experimental Figure 3. Material of ALBO-CA in the period of 15 days. On the microphotography there is experimental period of 15 days. In microphotography, a moderate thick capsule and a thin capsule and a mild inflammation reaction. mild vascular congestion and mild inflammation Lymphocytes and rare plasmocytes are present, which speaks in favor of a chronic inflammatory (H&E, ×400). reaction (H&E, ×200).

In control preparations after 15 days, the weakest inflammatory response was recorded in *ALBO*-CSHA control (grade 2), while control preparations for *ALBO*-CA were rated with mark 3 (moderate inflammatory response), and for MTA score 2.3 (mild to moderate inflammatory reaction). In all control preparations, a slight disturbance of the structure of the loose connective tissue (grade 2) was observed, followed by the formation of a thin fibrous capsule (grade 2). Vascular congestion was equally expressed in control group of *ALBO*-CSHA and MTA (grade 2.3), while in *ALBO*-CA control score was rated 2.6.

At the end of the evaluation period of 30 days, an identical inflammatory response of loose connective tissue at the site of implantation *ALBO*-CA, *ALBO*-CSHA and MTA (score 0.7) was recorded. The formation of thick or moderately thick fibrous capsules (Figure 5 and 6) was established, which was the thickest with *ALBO*-CA (grade 3.7). There were no statistically significant differences between the analyzed parameters.

In control preparations after 30 days, a weaker inflammatory response, lower vascular congestion, and better integrity of the binders were observed in control samples of *ALBO*-CSHA and MTA (grade 0.7), compared with *ALBO*-CA control (grade 1). A moderately thick or thick fibrous

capsule was formed (Figure 7), thicker in the control of MTA material (grade 3.2), while the *ALBO*-CA and *ALBO*-CSHA controls were rated 3.

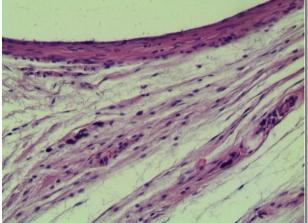
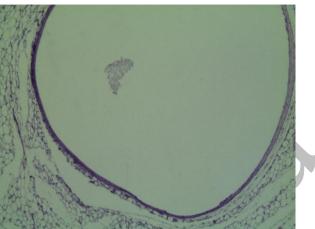


Figure 5. Material ALBO-CA in the experimental Figure 6. Material period of 30 days. The microphotography reveals a experimental period thick capsule and mild vascular congestion and mild microphotography, inflammation (H&E, ×400). subcutaneous fat tiss



ALBO-CSHA of in the period 30 days. In the of implantation the in subcutaneous fat tissue is encircled by a thick capsule without inflammatory response and congestion (H&E,×200).

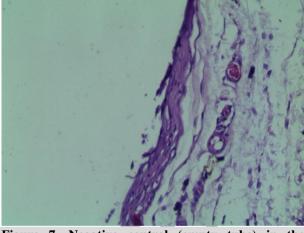


Figure 7. Negative control (empty tube) in the experimental period of 30 days. The microphotography reveals a thick capsule and a mild vascular congestion and mild inflammation (H&E, ×400).

DISCUSSION

Animal testing is a common method of checking new materials before their clinical examination.

There were almost no species and subspecies in the animal world that has not been used for the purposes of scientific research. However, when it comes to dental research, preference is given to rats. Although there are now hundreds of pure-blooded and about fifty crossed rat strains, the most commonly used in dental experiments are from Wistar and

Sprague-Dawley strains. The advantage in dental research for these experimental animals is primarily the similarity between their and human molars. In addition, rats can not vomit and are therefore used as a frequent starting model for numerous toxicity and carcinogenicity tests of dental materials. Of course, the advantage is their reasonable price and the possibility of simple securing. Like mice, rats have good reproductive skills and are well-grown in laboratory conditions [18].

Subcutaneous implants in rats are often used to evaluate the biological compatibility of various dental materials. The widespread use of this method comes from the fact that the implantation of the material into the subcutaneous tissue of the animal is accessible and simple, but also reliable in order to determine tissue irritation and the interaction between the tissue and the material itself.

[4,11,17]. Local reactions to the implanted implant can be quantitatively and qualitatively evaluated using various methods: histological analysis, scanning electron microscopy (SEM), transmision electro-microscopy (TEM), histochemical analysis. However, the most common method for studying the tissue compatibility of implanted material is histological analysis, and thickness of the fibrous capsule around the implant that is under the skin and used as an indicator of the biocompatibility of the material for decades [19].

Although a wide range of different types of tubes are in use today: dentine [20], silicone [21], teflon [3], polyethylene [22], polypropylene [23], our choice were polyethylene tubes, considering their inert nature and wide application. They do not cause a tissue reaction and in controlled and effective way expose tested material to the living tissue. They are easy to apply and sterilize [11,17,22].

Tissue around the tested materials showed the highest level of inflammation in first 7 days, with moderate damage to the structure of the connective tissue. The connective tissue around the MTA showed signs of the most severe inflammatory reaction evaluated as moderate and pronounced inflammation. This is in line with the findings of some other researchers [24,25] that a slightly more expressed initial inflammatory response to tissue implantation by MTA explain by high pH of this material, long-term follow-up of heat-trapping, as well as the stimulation of inflammatory cytokines.

In accordance with our results are also the results of the study *Opačić Galić V. et al.* [22], where 7 days after the subcutaneous implantation of the nanostructured CS-HA, CS and the control MTA, in rats of Wistar strain, the strongest inflammatory response was given by MTA (3.30 ± 0.48), while for CS and CS-HA it was graded with 3.00 ± 0.71 .

The results of this research showed the corelation between the strength of the inflammatory response and the thickness of the fibrous capsule. With time lag, a decrease in intensity of inflammation was noticed, with an increase in the thickness of the fibrous capsule in all tested materials. Experimental calcium aluminate cement (*ALBO*-CA) at the end of the observation period, after 30 days, caused an identical tissue reaction such as calcium silicate hydroxyapatite (*ALBO*-CSHA) and commercial mineral trioxide aggregate (MTA), with the best organized fibrous capsule around the material, which is probably due to the chemical nature and method of synthesis of this cement. The bioactivity of one material is also related to the method of its synthesis. The calcium aluminate nanostructured biomaterial, which was tested in this study, was produced by nanotechnology, by combining two methods: hydrothermal sol-gel method and the self-combusting wave methods. According to Chen et al. [26], the materials obtained by sol-gel processes are more bioactive than those obtained by other synthetic methods.

Garcia et al. [4] obtained biocompatibility results that fully correspond to the results of this study in the absence of inflammation and a significant increase in the thickness of fibrous capsule after 30 days of subcutaneous implantation of calcium aluminate cement (EndoBinder), calcium

silicate cement (Mineral trioxide aggregates) and calcium hydroxide in rats. Despite the fact that Endobinder releases a lower amount of calcium ions it caused a tissue reaction similar to that induced by MTA and calcium hydroxide in all observation periods. The authors explain this by the methodology applied in this in vivo study. Garcia et al. [4] concluded that from the biological point of view, EndoBinder became a promising option in endodontic therapy, but with the recommendation that additional research into the biological and physical-chemical properties of this new cement could be carried out before its application in the endodontic therapy of human populations.

Aminozarbian MG et al. [27] confirmed that the inflammatory response of the tissue to the implantation of CAAC calcium aluminate cement and the mixture of volostonite and calcium aluminate cement of WOLCA was, after 30 days, in rats comparable to MTA, which is in accordance with our findings , while CAAC Plus did not confirm biocompatibility as it induced a more intense inflammatory response than MTA. The authors explain this by the composition of CAAC Plus cement, with adding 5% Na-HMP dispersant in order to improve its properties, which on the other hand negatively affected biocompatibility.

Aguilar et al. [11] confirmed a better tissue reaction after calcium aluminate cement EndoBinder was compared to GMTA after 42 days of subcutaneous implantation of these materials in rat tissue. This is explained by the process of EndoBinder's synthesis, a low Ca content, which results in the release of a smaller amount of calcium ions, making it less irritant and cytotoxic to the tissues, without compromising its antimicrobial properties.

The reaction of the tissue to the empty tubes (negative control) in this experiment is similar to the findings of other researchers [19,28]. After seventh day, the most pronounced inflammatory response was confirmed, and this initial inflammatory reaction to the empty tubules was explained by the reaction to the surgical implantation procedure [19,28].

Results of the biocompatibility of *ALBO*-CSHA obtained in this study correspond to the findings of Petrovic et al. [17] and Saghiri et al. [29] In their studies at the end of the observation, there was no significant difference in the tissue inflammatory response to CS-HA and MTA, ie Bioagregate, nanostructured tricalcium silicate cement with the addition of calcium phosphate, and MTA after subcutaneous implantation in rats. The authors explain this finding by the fact that calcium-containing good biological properties owe a common ability to release calcium ions, and a similar reaction is expected in a situation where the new calcium silicate system comes into contact with tissue fluids. In the study of *Petrović et al.*, the application of a special synthesis method further favored the bioactivity of the examined CS-HA.

Contrary to this, Batur BY at al. [30] have come up with results that are inconsistent with the results of this study, as they confirmed statistically significantly better tissue response and a weaker inflammatory response of DiaRoot BioAggregate compared to MTA. As a possible reason for a better Bioaggregate result, the only fact is that their research was done *in vivo* conditions on

laboratory animals (Sprouge Dawley rats), as opposite to previous in vitro studies in which no statistically significant difference was found between MTA and BioAggregat.

CONCLUSION

The subcutaneous tissue of rats showed good tolerance of calcium aluminate nanostructured biomaterials and comparable to nanostructured calcium silicate *ALBO*-CSHA and commercial calcium silicate cement (MTA). The biocompatibility of this nanomaterial should be checked in other experimental studies before clinical screening on the human population.

NOTE

This paper is part of a research thesis entitled "Biocompatibility nanostructured biomaterials based on calcium aluminate" by Ognjenka Janković.

REFERENCES:

- 1. Modareszadeh MR, Chogle SA, Mickel AK, Jin G, Kowsar H, Salamat N, et al. Cytotoxicity of set polymer nanocomposite resin root-end filling materials. Int Endod J. 2011; 44: 154–61.
- Costaa CS, Heblingb J, Scheffelb DLS, Soaresc DGS, Bassoa FG, Ribeirod APD. Methods to evaluate and strategies to improve the biocompatibility of dental materials and operative techniques. Dent Mater. 2014; (30): 769–84.
- Eid AA, Niu LN, Primus CM, Opperman LA, Pashley DH, Watanabe I, et al. In Vitro Osteogenic/Dentinogenic Potential of an Experimental Calcium Aluminosilicate Cement. J Endod. 2013; 39(9): 1161–6.
- Garcia Lda FR, Huck C, Menezes de Oliveira L, de Souza PP, de Souza Costa CA. Biocompatibility of new calcium aluminate cement: tissue reaction and expression of inflammatory mediators and cytokines. J Endod. 2014; 40(12): 2024–9.
- 5. Garcia Lda FR. Calcium aluminate based-cements for endodontic application. J J Dent Res. 2014; 1(2): 008.
- 6. Van Dijken JW, Sunnegårdh-Grönberg K. A three year follow-up of posterior doxadent restorations. Swed Dent J. 2005; 29(2): 45–51.
- 7. Jefferies SR, Pameijer CH, Appleby DC, Boston D, Galbraith C, Loof J, et al. Prospective Observation of a New Bioactive Luting Cement: 2-Year Follow-Up. J Prosthodont. 2012; 21: 33–41.
- 8. Silva EJ, Herrera DR, RosaTP, Duque TM, Jacinto RC, Gomes BP, et al. Evaluation of cytotoxicity, antimicrobial activity and physicochemical properties of a calcium aluminate-based endodontic material. J Appl Oral Sci. 2014; 22(1): 61–7.
- 9. Pires-de-Souza Fde C, Moraes PC, Garcia Lda F, Aguilar FG, Watanabe E. Evaluation of pH, calcium ion release and antimicrobial activity of a new calcium aluminate cement. Braz Oral Res. 2013; 27(4): 324–30.
- 10. Garcia Lda FR, Rossetto HL, De Carvalho Panzeri F, Pires-de-Souza Fde. Shear bond strength of novel calcium aluminate-based cement (EndoBinder) to root dentine. Eur J Dent. 2014; 8(4): 498–503.
- 11. Aguilar FG, Garcia FR, Pires-de-Souza FCP. Biocompatibility of New Calcium Aluminate Cement (EndoBinder). J Endod. 2012; 38(3): 367–71.
- 12. Garcia LFR, Huck C, Scardueli CR, Costa CAS. Repair of Bone Defects Filled with New Calcium Aluminate Cement (EndoBinder). J Endod. 2015; 41(6): 1–7.
- 13. Wei W, Qi YP, Nikonov SY, Niu LN, Regina RLW, Mao J, et al. Effects of an Experimental Calcium Aluminosilicate Cement on the Viability of Murine Odontoblast-like Cells. J Endod. 2012; 38(7): 936–42.
- 14. Kramer PR, Woodmansey KF, White R, Primus CM, Opperman LA. Capping a Pulpotomy with Calcium Aluminosilicate Cement: Comparison to Mineral Trioxide Aggregates. J Endod. 2014; 40 (9): 1429–34.
- 15. de Oliveira IR, Ranieroa LJ, Leitea VMC, Castro-Raucci LMS, Oliveira PT, Pandolfellic VC. In vitro apatite-forming ability of calcium aluminate blends. Ceramics International. 2017; 43(13): 10071–9.
- 16. Liu WN, Chang J, Zhu YQ, Zhang M. Effect of tricalcium aluminate on the properties of tricalcium silicate-tricalcium aluminate mixtures: setting time, mechanical strength and biocompatibility. Int Endod J. 2011; 44(1): 41–50.

- Petrović V, Opačić Galić V, Jokanović V, Jovanović M, Jovanović BG, Živković S. Biocompatibility of a new nanomaterial based on calcium silicate implanted in subcutaneous connective tissue of rats. Acta Veterinaria-Belgrade. 2012; 62: 697–708.
- 18. Gregurić GG, Gračner D, Pavičić Ž. The behaviour of laboratory animals mice and rats. Veterinarska Stanica. 2015; 46 (3): 197–206.
- 19. Gomes-Filho JE, Watanabe S, Simonetti Lodi , Cintra LTA, Nery MJ, Filho JAO, et al. Rat tissue reaction to MTA FILLAPEX. Dent Traumatol. 2012; 28(6): 452–6.
- 20. Broon NJ, Martínez PAM, Muñiz GLR, Morales CT, Ángeles LHL, Bramante CM. Inflammatory response of Bioceramic material to dentin tube implantation in rats' subcutaneous tissue. Rev Odont Mex. 2016; 20(3): 174–8.
- Zmener O, Martinez Lalis R, Pameijer CH, Chaves C, Kokubu G, Grana D. Reaction of rat subcutaneous connective tissue to a mineral trioxide aggregate-based and a zinc oxide and eugenol sealer. J Endod. 2012; 38(9): 1233–8.
- Opačić-Galić V, Petrović V, Jokanović V, Živković S. Histological Evaluation of Tissue Reactions to Newly Synthetized Calcium Silicate- and Hydroxyapatite-Based Bioactive Materials: in vivo Study. Srp Arh Celok Lek. 2017; 145(7-8): 370–7.
- 23. Coleman NJ, Li Q. The Impact of Iodoform on the Hydration, Bioactivity and Antimicrobial Properties of White Portland Cement. MATEC Web of Conferences. 2017; 109: 04002.
- 24. Lotfi M, Vosoughhosseini S, Saghiri MA, Mesgariabbasi M, Ranjkesh B. Effect of white mineral trioxide aggregate mixed with disodium hydrogen phosphate on inflammatory cells. J Endod. 2009; 35(5): 703–5.
- 25. Camilleri J, Pitt Ford TR. Mineral trioxide aggregate: a review of the constituents and biological properties of the material. Int Endod J. 2006; 39(10): 747–54.
- Chen CC, Shie MY, Ding SJ. Human dental pulp cell responses to new calcium silicate-based endodontic materials. Int Endod J. 2011; 44: 836–42.
- 27. Aminozarbian MG, Barati M, Salehi I, Mousavi SB. Biocompatibility of mineral trioxide aggregate and three new endodontic cements: An animal study. Dent Res J. 2012; 9(1): 54–9.
- 28. Yavari HR, Shahi S, Rahimi S, Shakouie S, Roshangar L, Abbasi MM, et al. Connective tissue reaction to white and gray MTA mixed with distilled water or chlorhexidine in rats. Iranian Endod J. 2009; 4(1):25–30.
- Saghiri MA, Tanideh N, Garcia-Godoy F, Lotfi M, Karamifar K, Amanat D. Subcutaneous connective tissue reactions to various endodontic biomaterials: an animal study. J Dent Res Dent Clin Dent Prospects. 2013; 7(1): 15–21.
- Batur YB, Acar G, Yalcin Y, Dindar S, Sancakli H, Erdemir U. The cytotoxic evaluation of mineral trioxide aggregate and bioaggregate in the subcutaneous connective tissue of rats. Med Oral Patol Oral Cir Bucal. 2013; (4): 745–51.

DOI: https://doi.org/10.2298/SARH171211030J